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㉗ **Herbicide resistance in plants.**

㉘ This invention is directed to the production of plants, plant tissues and plant seeds which are resistant to inhibition by an herbicide which normally inhibits the growth and development of those plants, plant tissues and plant seeds. In particular this invention is directed to altered acetoxy-droxyacid synthase enzymes which are resistant to inhibition by herbicides which normally inhibit the activity of the synthase before such alteration. This invention further relates to genes encoding such enzymes, and to processes for utilizing these novel genes and enzymes. Further products of the invention include plants, plant tissues and seeds which exhibit resistance to such herbicides resulting from expression of genes encoding herbicide resistant acetoxy-droxyacid synthase enzyme.

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1. FIELD OF THE INVENTION

This invention relates to genes and enzymes which confer resistance to herbicides in plants, plant tissues and seeds. In particular, the invention involves
10 agronomically important crops which are resistant to herbicides, and which genetically transmit this characteristic to their progeny.

15

2. BACKGROUND OF THE INVENTION

2.1. WEED CONTROL

The use of herbicides for controlling weeds or plants in crops has become almost a universal practice.
20 The market for these herbicides approaches a billion dollars annually. Even with this extensive use, weed control remains a significant and costly problem for the farmer.

25

Present day herbicides used singly or in so-called tank mixes require good management to be effective. Time and method of application and stage of weed plant development are critical to getting good weed control with herbicides. Some weed species are simply
30 resistant to today's herbicides. Therefore, the production of effective herbicides increases in importance every year, especially as other weeds are controlled and thus reduce competition. Application of large amounts of marginally effective herbicides on these weeds can result
35 in a commitment to grow the same crop in subsequent years

because of chemical persistence in the soil which prevents rotation with a crop sensitive to that herbicide.

5 Other herbicides, while not used directly to
control weeds in field crops, are used as "total
vegetation control agents" to entirely eliminate weeds in
certain railroad and industrial situations. These
herbicides may be deposited on areas where crops are
10 planted by water run-off, or other natural means. Thus,
in fields affected by run-off from land on which total
vegetation control agents have been used, sensitive field
crops may be killed or their growth seriously inhibited.

15 Herbicides with greater potency, broader weed
spectrum and more rapid degradation in the soil would have
a significant impact on these problems. Unfortunately,
these compounds also have greater crop phytotoxicity.
Crop hybrids or varieties with resistance to the compounds
20 would provide an attractive solution by allowing the
compounds to be used without risk of damage to the crop.

2.2: TISSUE CULTURE OF MAIZE

25 Irrespective of the plant species, there are a
number of common features that apply to most tissue
culture programs. The technique of cell and tissue
culture has been widely developed, and much work has been
done on growth, metabolism and differentiation of tissue
culture of dicotyledons (Yamada, 1977, in Plant Cell,
30 Tissue and Organ Culture, eds. Reinert and Bajaj, pp.
144-159, Springer-Verlag, Berlin). However, successful
tissue culture studies with monocotyledons (e.g., the
cereal crops such as maize, rice wheat, barley, sorghum,
oats, rye and millet) leading to plant regeneration are
35 not as well documented as with dicotyledons. Success is

frequently dependent on choosing donor tissues for culture initiation which come from plants of appropriate genotype as well as physiological and development states. Other features which are obviously also important include the
5 organic and inorganic composition of the growth medium and the physical environment in which the cultures are grown.

In maize, the development of tissue cultures capable of plant regeneration was accomplished after the
10 identification of appropriate genotypes and donor tissues (Green and Rhodes, 1982 in Maize for Biological Research, ed. W.F. Sheridan, pp. 367-371, Plant Molecular Biology Association, Charlottesville, VA). The first method developed which regenerated plants from tissue cultures of
15 maize used immature embryos as donor tissues. With N6 or MS growth media (defined below in Section 6) and a synthetic auxin, such as 2,4-dichlorophenoxyacetic acid (2,4-D), tissue cultures develop rapidly from the scutellum of the embryos. The resulting cultures are
20 developmentally heterogeneous and contain a variety of tissue types. Removal of the 2,4-D from the growth medium permits these cultures to produce large numbers of regenerated plants. Cultures of this type have proved capable of regenerating plants for up to three years.

25 Another donor tissue from which regenerable tissue cultures of maize have been initiated are immature tassels. This tissue is the male flower and as it matures it is responsible for pollen production. Immature
30 embryos, inflorescences, and the few other tissues in cereals from which regenerating cultures have been initiated all have the common characteristic of juvenility. Regenerated plants obtained from tissue cultures are grown to maturity in a glasshouse, growth
35 chamber, or field. The progeny seed produced in crosses

with regenerated plants permits the evaluation of subsequent generations. The basic tissue culture methods developed for corn have been extended to many other cereal species.

5

An interesting development in recent years has been the occurrence of somatic embryogenesis in tissue cultures of maize. Somatic embryogenesis is the process where cells from callus, suspension, or protoplast

10 cultures develop into complete embryos similar to zygotic embryos produced in seeds. It is now possible to reliably initiate cultures of corn which have two important characteristics. One is that the callus cultures are friable, meaning that they are soft and loose in texture.

15 This property is important because cultures of this type exhibit rapid growth and it facilitates the initiation of suspension cell cultures. The other valuable attribute of these friable cultures is their ability to form very large numbers of somatic embryos. Microscopic examination

20 reveals the presence of many small, organized structures on the surface of the callus. These structures are young somatic embryos at various developmental stages. These friable cultures will retain their embryogenic potential for as long as two years and have shown the capacity to

25 produce extremely large numbers of somatic embryos.

The somatic embryos in these friable calli develop to maturity when the cultures are transferred to medium containing 5 to 6 percent sucrose and no hormones.

30 After approximately two weeks of growth on this medium, many embryos have become quite mature. They germinate rapidly and grow into plants when placed on MS or N6 medium containing 2% sucrose. The plants are then established in soil and are grown to maturity.

35

It is now well-documented that a high level of genetic variability can be recovered from plant tissue culture. It is well documented that spontaneous genetic variability in cultured plant cells may be the result of mutation (Meredith and Carlson, 1982, in Herbicide Resistance in Plants, eds. Lebaron and Gressel, pp. 275-291, John Wiley and Sons, NY). The frequency of mutants can also be increased by the use of chemical or physical mutagens. Some of this variability is of agronomic importance. Mutants for disease resistance have been obtained in sugarcane for Fiji disease, early and late blight in potato, and southern corn leaf blight in maize. In rice, maize, and wheat considerable variability for traits inherited as single genes of plant breeding interest have been recovered, including time of seed set and maturation, seed color and development, plant height, plant morphology, and fertility.

Tissue cultures of maize have been used to recover mutants for disease resistance and amino acid overproduction as described below.

Texas male sterile cytoplasm (cms-T) genotypes of maize are susceptible to the pathotoxin produced by the fungus Helminthosporium maydis race T while normal cytoplasm (N) genotypes are resistant (Gengenbach et al., 1977, Proc. Natl. Acad. Sci. USA 74: 5113-5117). Similarly, tissue cultures obtained from cms-T genotypes are susceptible to the pathotoxin while N genotype cultures are resistant. The pathotoxin from H. maydis race T was used to select resistant cell lines from susceptible cms-T cultures using a sublethal enrichment selection procedure. After five cycles of increasing selection pressure, cell lines were recovered which were resistant to lethal levels of the pathotoxin. Plants

regenerated from these cell lines also were resistant to the pathotoxin and were male-fertile. Genetic analysis of progeny obtained from resistant, male-fertile plants showed that both traits were maternally inherited.

- 5 Infection of plants with H. maydis race T spores demonstrated that selection for pathotoxin resistance also resulted in resistance to the disease organism by plants.

Selection for resistance to growth inhibition by
10 lysine plus threonine in equimolar concentrations (LT) in tissue cultures of maize yielded a stable resistant line, LT19 (Hibberd and Green, 1982, Proc. Natl. Acad. Sci. USA 79: 559-563). Genetic analysis of progeny of plants regenerated from LT19 showed that LT resistance was
15 inherited as a single dominant nuclear gene. Tissue cultures initiated from resistant embryos required 5-10 times higher levels of LT to inhibit growth than did cultures from LT-sensitive embryos. LT resistance in LT19 was expressed as reduced sensitivity of root and shoot
20 growth to the presence of LT. The free pool of threonine was increased 6 times in cultures initiated from immature embryos of LT-resistant plants, and 75-100 times in kernels homozygous for LT19, as compared to cultures and kernels from LT-sensitive embryos and plants,
25 respectively. Overproduction of free threonine increased the total threonine content in homozygous LT19 kernels by 33-59%. The results demonstrate that LT resistance selected with tissue culture methods was heritable and was expressed in cultures, seedlings, and kernels.

30

2.3. MECHANISMS OF HERBICIDE RESISTANCE

There are three general mechanisms by which plants may be resistant to, or tolerant of, herbicides.

- 35 These mechanisms include insensitivity at the site of

action of the herbicide (usually an enzyme), rapid metabolism (conjugation or degradation) of the herbicide, or poor uptake and translocation of the herbicide.

Altering the herbicide site of action from a sensitive to
5 an insensitive form is the preferred method of conferring resistance on a sensitive plant species. This is because resistance of this nature is likely to be a dominant trait encoded by a single gene and is likely to encompass whole families of compounds that share a single site of action,
10 not just individual chemicals. Therefore, detailed information concerning the biochemical site and mechanism of herbicide action is of great importance and can be applied in two ways. First, the information can be used to develop cell selection strategies for the efficient
15 identification and isolation of appropriate herbicide resistant variants. Second, it is used to characterize the variant cell lines and regenerated plants that result from the selections.

20

2.4. HERBICIDE RESISTANCE SELECTION

Tissue culture methods have been used to select for resistance (or tolerance) using a variety of herbicides and plant species (see review by Meredith and
25 Carlson, 1982, in Herbicide Resistance in Plants, eds. Lebaron and Gressel, pp. 275-291, John Wiley and Sons, NY). The results of these investigations can be separated into two categories based on whether or not herbicide tolerance was stably inherited and expressed in the
30 progeny of plants regenerated from the selected resistant cultures. This criterion clearly establishes the mutant nature of the selected trait. A number of tissue culture studies have been conducted to select for tolerance to 2,4-dichlorophenoxyacetic acid (2,4-D) in carrot, tobacco
35 and white clover, to amitrole in tobacco, to asulam in

celery, and to paraquat in tobacco, in none of which was the mutant nature of the resistance trait established by genetic analysis. These studies have therefore provided little evidence demonstrating the feasibility of tissue culture selection methods to produce herbicide resistant mutant plants which transmit the trait to progeny which express the resistance.

Three studies are available, however, which provide evidence that tissue culture methods can be utilized to obtain herbicide resistant mutants. Tobacco selected for tolerance to bentazon and phenmedipham yielded resistant plants (Radin and Carlson, 1978, Genet. Res., Camb., 32: 85-90). Genetic analysis of progeny from regenerated plants yielded data in the F2 generation confirming a genetic basis for resistance in 8 bentazon and 2 phenmedipham selected lines. The F2 segregation ratios indicated single gene recessive mutations for most of the lines except two bentazon lines in which two genes were indicated.

Chaleff and Parsons (1978, Proc. Natl. Acad. Sci. USA 75: 5104-5107) used tissue culture selection methods to isolate picloram resistant mutants from tobacco suspension cultures. Plants were regenerated from six of seven resistant lines selected. Resistance to picloram was transmitted to progeny in four of these lines and was expressed in both plants and callus tissues. In all four cases, segregation ratios were those expected from dominant single-gene mutations. In additional genetic analysis two of these mutants were shown to be linked.

Tomato callus lines were selected for the ability to grow at paraquat concentrations lethal to wild-type cells (Thomas and Pratt, 1983, Theor. Appl. Genet.

63: 109-113). Diploid plants were regenerated from 9 of the 19 paraquat resistant callus lines isolated. New callus cultures were initiated from these regenerated plants and typically showed at least a 30 fold increase
5 over wild-type in resistance to paraquat. Tests on callus lines initiated from sexual progeny of regenerated plants showed that the paraquat resistance phenotype of three lines resulted from dominant nuclear mutations. Paraquat spray experiments indicated that slight paraquat
10 resistance was expressed at the plant level in only one of the resistant lines.

2.5. HERBICIDAL IMIDAZOLINONES

15 A broad selection of imidazolinones, particularly 2-(2-imidazolin-2-yl)pyridines and 2-(2-imidazolin-2-yl)quinolines, or derivatives thereof, exhibit herbicidal activity. See, for example, European Patent Application 81 103638.1 naming Los as inventor and American Cyanamid
20 Company as applicant.

Exemplary herbicides of particular interest described in this application are 2-(5-isopropyl-5-methyl-4-oxo-2-imidazolin-2-yl)nicotinic acid (AC 243,997), 2-(5-isopropyl-5-methyl-4-oxo-2-
25 imidazolin-2-yl)3-quinolinecarboxylic acid (AC 252,214), [5-ethyl, 2-(5-isopropyl-5-methyl-4-oxo-2-imidazolin-2-yl)nicotinic acid (AC 263,499), and acid addition salts thereof.

30 For purposes of reference in the present specification, the herbicides described in this Section 2.5, and structurally related herbicidal compounds, are collectively referred to as imidazolinones or the imidazolinone family of herbicides.

2.6. HERBICIDAL SULFONAMIDES

Certain sulfonamides exhibit general and selective herbicidal activity against plants. Such herbicidal sulfonamides are disclosed in at least the following issued United States Patents:

4,435,206	4,370,480	4,302,241
4,424,073	4,370,479	4,293,330
4,417,917	4,369,320	4,257,802
4,398,939	4,369,058	4,231,784
4,394,506	4,368,067	4,225,337
4,391,627	4,348,219	4,221,585
4,383,113	4,342,587	4,214,890
4,378,991	4,339,267	4,190,432
4,372,778	4,339,266	4,169,719
4,371,391	4,310,346	4,127,405

15

One such herbicidal sulfonamide of particular interest is 2-chloro-N-[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide, also known as chlorsulfuron.

For purposes of reference in the present specification, the herbicides referred to in this Section 2.6, and structurally related herbicidal compounds, are collectively referred to as herbicidal sulfonamides.

3. SUMMARY OF THE INVENTION

30

This invention is directed to agronomically important plants, plant tissues, and plant seeds which are resistant to inhibition by an herbicide at concentrations which normally inhibits the growth and development of those plants, plant tissues and plant seeds. The present invention contemplates the introduction of herbicide

resistance into any agronomically important crop including, but not limited to certain dicotyledons, monocotyledons, and specifically cereal crops such as rice, maize, wheat, barley, sorghum, oats, rye, millet and the like.

In particular this invention is directed to processes for the production of plants, plant tissues, and plant seeds which contain an enzyme which is resistant to inhibition by an herbicide at a concentration which normally inhibits the activity of this enzyme before alteration. This enzyme, an altered acetohydroxyacid synthase (AHAS), confers resistance to certain herbicides in plants, plant tissues and seeds. This invention is also directed to processes and compositions for making and utilizing this altered enzyme, as well as the gene encoding the resistant enzyme. One particular embodiment of the present invention is directed to the production of an herbicide resistant cell line possessing an alteration at the primary site of action of an herbicide. The present invention utilizes cell culture technology to isolate, characterize and develop herbicide resistant maize lines which genetically transmit this herbicide resistance characteristic to their progeny.

The present invention also provides a method of generating plants, plant tissues and seeds from these resistant cell lines which contain a gene coding for an altered acetohydroxyacid synthase resistant to inhibition by an herbicide at a concentration which normally inhibits the activity of this enzyme before alteration.

In the examples presented herein, a novel enzyme, an altered acetohydroxyacid synthase which is resistant to inhibition by certain imidazolinone and/or sulfonamide

herbicides (at concentrations which normally inhibit the activity of the unaltered enzyme) is described. This enzyme confers resistance to the herbicides in plants, plant tissues and seeds. Maize genotypes expressing this
5 altered AHAS are described. This maize genotype may be used with the imidazolinone or sulfonamide herbicides to effectively combat weed problems in maize production.

The present invention further contemplates that
10 herbicide resistance to the compounds described herein may be conferred by a variety of other mechanisms. For example, resistance may be conferred by a gene coding for an enzyme which may be obtained from any source including, but not limited to eukaryotic organisms, procaryotic
15 organisms, or it may be made in whole or part by chemical or enzymatic synthetic methods. Other mechanisms of resistance are discussed in Section 2.3 supra.

It is to be understood that the following
20 detailed description presents a single embodiment of the invention. This embodiment relates to an alteration in a particular enzyme, acetohydroxyacid synthase, which renders plants, plant tissues and seeds resistant to certain imidazolinones, the herbicides for which
25 resistance was selected. Unexpectedly, these plants, plant tissues and seeds were also found resistant to certain sulfonamide herbicides. Thus, the altered enzyme disclosed herein may confer resistance to a variety of herbicides which inhibit acetohydroxyacid synthase as
30 their primary site of action.

4. BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 shows the growth of maize cell suspension
35 cultures as a function of AC 243,997 concentration with

(open circles) or without (closed circles) media supplementation with 1.0 mM each of leucine, valine and isoleucine.

5 FIG. 2 shows the biosynthetic pathway for production of leucine, valine and isoleucine in plants.

FIG. 3 shows the shoot length growth of seedlings grown in the presence or absence of AC 252,214.

10

FIG. 4 shows the root length growth of seedlings grown in the presence or absence of AC 252,214.

5. DETAILED DESCRIPTION OF ONE EMBODIMENT OF THE INVENTION

15

This embodiment of the invention relates to plants, plant tissues, and plant seeds which contain an enzyme which is resistant to inhibition by imidazolinone and/or sulfonamide herbicides. The enzyme is an altered
20 acetohydroxyacid synthase which confers resistance to the above-mentioned herbicides in plants, plant tissues and seeds. Methods and compositions are provided for producing plants, plant tissues and seeds containing a gene coding for an altered AHAS. Also described are cell
25 culture selection techniques to select for novel maize genotypes resistant to certain imidazolinone and/or sulfonamide herbicides. The production of these maize lines encompasses isolation, characterization, and development of these maize lines and regeneration of
30 plants from these cultures which are resistant to the herbicides.

The method of this invention may be divided into the following areas for the purposes of description:

35 (1) determination of the primary site of action of the

imidazolinone family of herbicides; (2) characterization of the affects of the imidazolinone herbicides on maize cell cultures and the strategy for selection of herbicide resistant cell lines; (3) selection and characterization of herbicide resistant cell lines; (4) regeneration of herbicide resistant plants and production of seed; and (5) development of herbicide resistant commercial hybrid seed.

5.1. DETERMINATION OF THE PRIMARY SITE OF ACTION OF THE IMIDAZOLINONE FAMILY OF HERBICIDES

10

The biochemical site of action of the herbicides, including but not limited to the imidazolinone family and sulfonamide family of herbicides, in maize tissue is determined by first evaluating which general cell metabolic processes are affected by tissue exposure to the phytotoxic compounds. The specific site of action is then localized by in vitro evaluation of the individual reactions within the affected metabolic pathway or process.

20

The primary site of action may be determined by adding to tissue exposed to herbicide various products of the pathway(s) which is suspected to be affected by the herbicide. Those products which reverse the growth inhibitory effects of the herbicide indicate the pathway(s) affected by the herbicides.

25

5.2. CHARACTERIZATION OF THE EFFECTS OF THE IMIDAZOLINONE HERBICIDES ON PLANT CELL CULTURES AND STRATEGY FOR SELECTION OF HERBICIDE RESISTANT CELL LINES

30

Efficient selection of a desired herbicide resistant mutant using tissue culture techniques requires careful determination of selection conditions. These conditions are optimized to allow growth and accumulation

35

of rare herbicide resistant cells in the culture while inhibiting the growth of the bulk of the cell population. The situation is complicated by the fact that the vitality of individual cells in a population is highly dependent on
5 the vitality of neighboring cells.

Conditions under which cell cultures are exposed to the herbicides are determined by the characteristics of the interaction of the compounds with the tissue. Such
10 factors as the accumulation of the compounds by cells in culture, and the persistence and stability of the compounds, both in the media and in the cells, need to be considered. Also important is whether the effects of the compounds can be readily reversed following their removal.

15

Aside from factors associated with the chemistry of the herbicidal compounds, their effects on culture viability and morphology need to be carefully evaluated. It is especially important to choose herbicide exposure
20 conditions which have no impact on plant regeneration capability of cultures. Choice of herbicide exposure conditions is also influenced by whether the herbicide kills cells or simply inhibits cell divisions.

25

The choice of a selection protocol is dependent upon the considerations described supra. Either of the protocols briefly described below may be utilized in the selection procedure, although the present invention is not limited to these procedures. In the first protocol,
30 finely divided cells in liquid suspension culture are exposed to high herbicide levels for brief periods of time. Surviving cells are then allowed to recover and accumulate and are then reexposed for subsequently longer periods of time. Alternatively, organized, partially
35 differentiated cell cultures are grown and subcultured

with continuous exposure to initially low herbicide levels. Herbicide concentrations are then gradually increased over several subculture intervals.

5.3. SELECTION AND CHARACTERIZATION
OF RESISTANT CELL LINES

5

Selections are carried out until cells or tissue are recovered which are observed to be growing well in the presence of normally toxic herbicide levels. These cell
10 "lines" are then repeatedly subcultured in the presence of herbicide and characterized. The amount of resistance which has been obtained is determined by comparing the growth of these cell lines with the growth of unselected cells or tissue in the presence of various herbicide
15 concentrations. Stability of the herbicide resistance trait of the cultured cells may be evaluated by simply growing the selected cell lines in the absence of herbicide for various periods of time and then analyzing growth after reexposing the tissue to herbicide.

20

In the present invention, cell lines which are resistant by virtue of having an altered herbicide site of action are of primary interest. Cell lines may also be tested for resistance to herbicides structurally related
25 to the selection agent. The resistant cell lines may also be evaluated using in vitro chemical studies to verify that the site of action of the herbicide is altered to a form which is less sensitive.

30

5.4. PLANT REGENERATION AND PRODUCTION OF SEED

Cell lines exhibiting satisfactory levels of resistance by virtue of having an altered herbicide site
of action are put through a plant regeneration protocol to
35 obtain mature plants and seeds expressing the resistance

trait. The plant regeneration protocol allows the development of somatic embryos and the subsequent growth of roots and shoots. To determine that the herbicide resistance trait is expressed in differentiated organs of the plant and not solely in undifferentiated cell culture, early plant regeneration steps are carried out in the presence of herbicide levels that will normally inhibit shoot and root formation and growth.

10 Mature plants are then obtained from cell lines that are known to express the trait. If possible, the regenerated plants are self pollinated. Otherwise pollen obtained from the regenerated plants is crossed to seed grown plants of agronomically important inbred lines. 15 Conversely, pollen from plants of these inbred lines is used to pollinate regenerated plants. The genetics of the trait is then characterized by evaluating the segregation of the trait in first and later generation progeny. The heritability and expression in plants of traits selected in tissue culture are of particular importance if the 20 traits are going to be commercially useful.

5.5. DEVELOPMENT OF HERBICIDE RESISTANT COMMERCIAL HYBRID SEED

25 Seed from plants regenerated from tissue culture is grown in the field and self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines which are evaluated for herbicide resistance in the field under a range of environmental 30 conditions. Herbicide resistance must be sufficient to protect the corn at the maximum labeled rate under field conditions which cause the herbicide to be most active. Appropriate herbicide concentrations and methods of application are those which have been developed for the 35 herbicide in question.

The commercial value of herbicide resistant corn is greatest if many different hybrid combinations with resistance are available for sale. The farmer typically grows more than one kind of hybrid based on such differences as maturity, standability or other agronomic traits. Additionally, hybrids adapted to one part of the corn belt are not adapted to another part because of differences in such traits as maturity, disease and insect resistance. Because of this, it is necessary to breed herbicide resistance into a large number of parental lines so that many hybrid combinations can be produced.

Adding herbicide resistance to agronomically elite lines is most efficiently accomplished if the genetic control of herbicide resistance is understood. This requires crossing resistant and sensitive plants and studying the pattern of inheritance in segregating generations to ascertain whether the trait is expressed as a dominant or recessive, the number of genes involved, and any possible interaction between genes if more than one are required for expression. This genetic analysis can be part of the initial efforts to convert agronomically elite, yet sensitive, lines to resistant lines.

A conversion process (backcrossing) is carried out by crossing the original resistant line by sensitive elite lines and crossing the progeny back to the sensitive parent. The progeny from this cross will segregate such that some plants carry the resistance gene (S) whereas some do not. Plants carrying the resistance gene (S) will be crossed again to the sensitive parent resulting in progeny which segregate for resistance and sensitivity once more. This is repeated until the original sensitive parent has been converted to resistant line, yet possesses all other important attributes as originally found in the

sensitive parent. A separate backcrossing program is implemented for every sensitive elite line that is to be converted to an herbicide resistant line.

5 Subsequent to the backcrossing, the new resistant lines and the appropriate combinations of lines which make good commercial hybrids are evaluated for herbicide resistance, as well as a battery of important agronomic traits. Resistant lines and hybrids are produced which
10 are true to type of the original sensitive lines and hybrids. This requires evaluation under a range of environmental conditions where the lines or hybrids will generally be grown commercially. Parental lines of hybrids that perform satisfactorily are increased and used
15 for hybrid production using standard hybrid seed corn production practices.

5.6. ALTERNATE METHODS OF OBTAINING
HERBICIDE RESISTANT MUTANTS

20 Generally, any alteration or replacement of AHAS which leads to herbicide resistance in tissue culture, seed and regenerated plants may be utilized in this embodiment of the present invention. AHAS may be altered
25 or replaced in any plant species; of especially great importance are the agronomic and horticulture crops in which herbicides are utilized. Such plants include for example, monocotyledenous plants and cereal crops such as maize, rice, wheat, barley, sorghum, oats, rye and the
30 like. The alteration of AHAS may be accomplished by any of a variety of means, including but not limited to the following methods: (1) spontaneous variation and direct mutant selection in tissue cultures; (2) direct or
indirect mutagenesis procedures on tissue cultures of all
35 types, seeds and plants; and (3) isolation of genes,

manipulation, modification, or synthesis in whole or part of genes using molecular biology, chemical technologies, and state-of-the-art procedures and reintroduction of herbicide resistance genes into plants.

5

Additionally, any type of AHAS modification which leads to a change in resistance to, or tolerance of, chemical compounds applied to plants may be utilized. These changes may include alterations in enzyme structure and changes in enzyme expression and/or function.
10 Chemical compounds include not only those which may be synthesized by techniques of organic chemistry, but also naturally occurring compounds which may affect AHAS activity in the plant, such as leucine and valine.
15 Herbicide resistance may also be accomplished by replacement or supplementation (i.e., gene therapy or the addition of extra genes), by any means, of an endogenous AHAS with any other AHAS from another source, including but not limited to prokaryotic or eukaryotic organisms or
20 by a total or partial chemical synthesis of a gene that catalyzes the same reactions as AHAS.

Genes encoding AHAS are common to the aspartate and branched chain amino acid pathways in plants and microorganisms (Umbarger, 1978, Ann. Rev. Biochem.
25 47: 533-606). AHAS catalyzes the first step in the synthesis of the amino acids leucine, valine and isoleucine. AHAS activity is regulated in vivo by various end products or combinations of end products of the biosynthetic pathway. AHAS genes with a variety of
30 regulatory characteristics are available from microorganisms including, but not limited to, Escherichia coli, yeast and Neurospora. These might include AHAS genes which are naturally resistant to the imidazolinone herbicides. For example, the isozymes of
35

AHAS (I, II and III) present in the bacterium E. coli are insensitive to millimolar concentrations of AC 243,997 when assayed in vitro. Resistance of E. coli to AC 243,997 therefore appears to be the result of an
5 insensitive site of action as opposed to being a result of metabolism of the herbicide. Resistance in other organisms is likely to be of the same nature, providing a variety of sources of a gene encoding an herbicide resistant AHAS. The genes encoding all three E. coli
10 isozymes have been cloned and sequenced. Selection for imidazolinone resistant mutant AHAS genes in microorganisms or plants including plant tissue cultures would provide a diverse source of genes which, when transferred to plants, produce herbicide resistant plants.

15

To introduce isolated genes or groups of genes into the genome of plant cells an efficient host gene vector system is necessary. The foreign genes should be expressed in the transformed plant cells and stably
20 transmitted (somatically and sexually) to the next generation of cells produced. The vector should be capable of introducing, maintaining and expressing a gene in plant cells, from a variety of sources, including but not limited to plants, animals, bacteria, fungi or
25 viruses. Additionally it should be possible to introduce the vector into a wide variety of plants. The location of the new gene in the plant genome may be important in determining effective gene expression of the genetically engineered plant. In addition, to be effective, the new
30 gene must be passed on to progeny by normal breeding.

Directed genetic modification and expression of foreign genes in dicotyledonous (broad-leaved) plants such as tobacco, potato and alfalfa has been shown to be
35 possible using the T-DNA of the tumor-inducing (Ti)

plasmid of Agrobacterium tumefaciens. Using recombinant DNA techniques and bacterial genetics, any foreign piece of DNA can be inserted into T-DNA in Agrobacterium. Following infection by the bacterium or Ti plasmid, the
5 foreign DNA is inserted into the host plant chromosomes, thus producing a genetically engineered cell and eventually a genetically engineered plant. A second approach is to introduce root-inducing (Ri) plasmids as the gene vectors. While Agrobacterium appear to attack
10 only dicots, many important crop plants (corn, wheat, rice, barley, oats, sorghum, millet and rye) are monocots and are not known to be susceptible to transformation by Agrobacterium. The Ti plasmid, however, may be
15 manipulated in the future to act as a vector for monocot plants. Additionally, using the Ti plasmid as a model system it may be possible to artificially construct gene vectors for monocot plants.

Ti-plasmids or other plasmids might also be
20 introduced into monocots by artificial methods such as microinjection, or fusion between monocot protoplasts and bacterial spheroplasts containing the T-region which can then be integrated into the plant nuclear DNA.

Genetic engineering of plants may be accomplished
25 by introducing the desired DNA containing functional genes encoding herbicide insensitive AHAS enzymes into plant tissues or cells using DNA molecules of a variety of forms and origins including but not limited to: plant pathogens
30 such as DNA viruses like Cauliflower Mosaic virus (CaMV) or geminiviruses, RNA viruses, and viroids; DNA molecules derived from unstable plant genome components like extrachromosomal DNA elements in organelles (e.g.,
chloroplasts or mitochondria), or nuclearly encoded
35 controlling elements; DNA molecules from stable plant

genome components (e.g., origins of replication and other DNA sequences which allow introduced DNA to integrate into the organellar or nuclear genomes and to replicate normally, to segregate normally during cell division and sexual reproduction of the plant and to be inherited in succeeding generations of plants).

The Cauliflower Mosaic virus (CaMV) has a gene the function of which is to prevent certain insects from destroying the virus. The remaining parts of the gene are redundant and can be replaced with a gene(s) which is useful to the plant breeder. The geminiviruses (or twin viruses) which are composed of two strands of DNA wrapped up in twin capsules of proteins can be used to transfer foreign genes into monocot plants. Transposons may also be used to carry foreign genes into plant DNA.

DNA containing AHAS genes may be delivered into the plant cells or tissues directly by infectious plasmids, such as Ti, viruses or microorganisms like A. tumefaciens, the use of liposomes, microinjection by mechanical or laser beam methods, by whole chromosome fragments, and by direct spraying of an infectious plant virus on crops.

Herbicide resistant plants can be developed using any of the means described supra and also other means including, but not limited to, conventional genetic and plant breeding procedures, whole plant genetic methods and somatic hybridization by protoplast fusion.

6. EXAMPLE

According to one particular embodiment of the present invention, the mechanism of action of the

imidazolinone herbicides and the primary site of action of herbicidal activity was determined to be the enzyme acetohydroxyacid synthase (AHAS). Maize tissue cultures resistant to an imidazolinone herbicide were selected and 5 plants which were resistant to the herbicide were regenerated from these cultures. An altered AHAS enzyme was identified in herbicide resistant cultures. A detailed description of the invention is presented in the subsections below.

10

6.1. HERBICIDE SITE OF ACTION

Elucidation of the biochemical mechanism of action of the imidazolinone family of herbicides was carried out using the herbicidal compound AC 243,997 15 (2-(5-isopropyl-5-methyl-4-oxo-2-imidazolin-2-yl) nicotinic acid) hereinafter referred to as 997 (American Cyanamid Co., Princeton, NJ). This compound is phytotoxic to most, if not all, higher plant species.

20

6.1.1. PLANT MATERIAL

Nonregenerable Black Mexican Sweet Corn cell suspension cultures (i.e., cell cultures which lack the 25 ability to regenerate plants) were obtained from Dr. Burle Gengenbach, Department of Agronomy and Plant Genetics, University of Minnesota, St. Paul, MN. Stock cell cultures were maintained in liquid Murashige and Skoog (MS) medium (Murashige and Skoog, 1962, *Physiol. Plant* 30 15: 473) containing 2 mg of 2,4-dichlorophenoxyacetic acid (2,4-D) per liter, and subcultured weekly by diluting to 1/10 the original volume with fresh medium. Cultures were grown in the dark at 26°C in 500 ml flasks containing 125 ml media.

35

6.1.2. GROWTH INHIBITORY EFFECTS OF 997

Homogeneous, undifferentiated plant cell cultures (i.e., cell suspension cultures) provide a convenient system for studying herbicide mechanism of action at the biochemical level. An initial comparison of growth inhibitory effects of 997 on maize cell suspension cultures and maize seedlings derived from immature embryos was made. Growth inhibitory studies indicated that maize cell suspension cultures and maize seedlings responded in a similar way to 997, therefore maize cell suspension cultures were used as a model system in all studies of the biochemistry of herbicide action in the present embodiment of this invention.

Growth inhibition studies were carried out in 125 ml flasks containing 30 ml of test media. 997 was filter sterilized and added to cooling media following autoclaving. Test media were inoculated with cells to give a settled cell volume (15 minutes without centrifugation) of 0.2 ml per 10 ml of suspension. After either six or seven days growth settled cell volumes were again quantitated.

The herbicidal effects of 997 on maize seedlings were determined by evaluating germination and seedling growth from excised, mature maize embryos (Green, 1974, Crop Sci. 14: 54-58). Seeds were sterilized for 10 minutes in 0.5% hypochlorite and then imbibed for 24 hours in sterile, distilled water. Mature embryos were aseptically excised and plated on solid MS media (9% agar) containing various concentrations of 997 and no 2,4-D. Embryos were placed with their shoot axis away from the media. Shoot growth was quantitated after seven days.

Growth of both seedlings and suspension cultures was 50% inhibited at extremely low concentrations of herbicide, i.e., 1 to 3×10^{-8} M in the media. Six maize inbred lines with different backgrounds, Al88, A632 (Minnesota Crop Improvement Assoc., St. Paul, MN); W117 (University of Wisconsin, Madison, WI); B37, B73 (Iowa Crop Improvement Assoc.); and Mol7 (Illinois Crop Improvement Assoc., Champaign, IL) were tested and found to have identical sensitivities to 997 within the experimental limits of the assay. The highly differentiated seedlings and the undifferentiated suspension cells both showed a high degree of sensitivity to the herbicide which suggested that plant development was not required for expression of sensitivity, and the suspension cells could be used for preliminary studies on determining the mechanism of action of the compound 997.

6.1.3. EFFECTS OF 997 ON PROTEIN SYNTHESIS

Experiments were conducted to determine whether growth inhibitory levels of 997 had inhibitory effects on cell protein synthesis.

Suspension culture cells were preincubated for 48 hours in the presence or absence of 1×10^{-5} M 997 and then exposed for 12 hours to 75 μ Ci of either 35 S-methionine (1210 Ci/mmol, New England Nuclear, Boston, MA) or 3 H-DL-leucine (55 Ci/mmol, ICN Pharmaceuticals Inc. - K and K Laboratories, Plainview, NY) per 0.1 ml of cells at a cell density of 10^6 volume cells/volume suspension. Cells were then centrifuged 300 x g, washed with cold MS culture media, and ground with a cold mortar and pestle in the presence of one volume of 6M urea. The resulting extract was centrifuged at 12,800 x g for 5 minutes. The pellet was then reextracted and, following centrifugation, the extracts

were combined. Protein was precipitated by the addition of 10% w/v trichloroacetic acid (TCA). Protein was removed by centrifugation and resolubilized in 6M urea. Incorporation of radiolabel into protein was determined by 5 scintillation counting.

At a concentration of 1×10^{-5} M, 997 was found to significantly inhibit protein synthesis as determined by quantitating incorporation of radiolabeled amino acids 10 into TCA-precipitable cell components. In experiments with cells at different post-subculture intervals, protein synthesis inhibition ranged from 50% to 77% on a cell fresh weight basis.

15 For determination of whether herbicide effects on protein synthesis were nonspecific, proteins were separated electrophoretically in one dimension on SDS (sodium dodecyl sulfate) 10% polyacrylamide slab gels (Laemmli, 1970, Nature 227: 680) and incorporation of 20 radiolabel visualized by fluorography (data not shown). Cell volumes of 0.4 ml were exposed to ^{35}S -methionine in the presence or absence of 1×10^{-6} M herbicide as described supra, ground with mortar and pestle, and then 25 extracted with 0.75 ml SDS sample buffer. The extracts were clarified by centrifugation and 20 ul or 4 ul samples loaded directly in the gels. The inhibition appeared to be nonspecific in nature. All major 6 M urea soluble intracellular protein components showed decreased levels of radiolabel.

30

6.1.4. DETERMINATION OF THE MECHANISM OF PROTEIN SYNTHESIS INHIBITION AND ITS RELATIONSHIP TO GROWTH INHIBITION

35 The effect of 997 on pool sizes of free amino acids was analyzed to determine if alterations in amino

acid metabolism were associated with the changes in protein synthesis (See Table 1).

TABLE 1

Free Amino Acid Levels in Presence
and Absence of 1×10^{-5} M 997
(nmol per g fresh weight tissue)
Corn in Suspension Cultures

	Amino Acid	Control	997 Treatment	
			3.5 hr.	48 hr.
	arg	52	60 ^a	55
	trp	22	30 ^a	30
15	lys	150	212	240
	his	146	184	310
	phe	124	151	195
	tyr	96	112	130
20	leu	196	120	16
	ile	121	120	38
	met	51	100	405
	val	397	355	48
	cystine ^b	18	103	435
25	ala	336	600 ^a	455
	gly	76	98	60
	glu	235	360	175
	ser	378	540	1160
	thr	69	89	N.D.
30	asp	193	175	100

^a Estimated

^b Oxidized cysteine

N.D. = not done

Suspension culture cells were treated for 3.5 hours or 48 hours with 1×10^{-5} M 997. Cells were removed from suspension by filtration and 1 gram fresh weight quantities were ground with sand with a mortar and pestle in 1 ml 5% TCA. Four ml of 5% TCA were added and the extract centrifuged at $10,000 \times g$ for 10 minutes. The pellet was reextracted and the supernatants combined and subjected to cation exchange chromatography. The samples were applied to Bond Elut SCX columns (Analytichem International Inc., Harbor City, CA) which had been equilibrated with 5% TCA. Following sample application, the columns were washed with 5 ml distilled, deionized water, and the amino acids were eluted with 2 ml aliquots of 5% v/v triethylamine. The aliquots were combined (6 ml), lyophilized, and subjected to amino acid analysis with a Kontron Liquemat III amino acid analyzer (Hesber, Middlesex TW5 0QU, England).

997 had a differential effect on the free amino acid levels in corn tissue growing in culture. The total free amino acids levels in Black Mexican Sweet Corn tissue treated with 997 increased over control cultures (Table 1). The biosynthetically related amino acids, serine, methionine, and cysteine, underwent the greatest increases during herbicide exposure (3, 8, and 24 fold, respectively, over a period of 48 hours). However, the biosynthetically related amino acids, leucine, valine, and isoleucine, decreased after herbicide treatment. After 48 hours of herbicide exposure, the levels of these amino acids decreased to 8, 12 and 31%, respectively, the levels seen in untreated cell cultures.

Cells were tested to determine whether the herbicide induced inhibition of growth was the result of the decreased availability of these three branched chain

amino acids. Cells were exposed to toxic levels of 997
(1×10^{-6} M) in media either supplemented with 1 mM
concentrations of leucine, valine, and isoleucine or with
no supplementation. Culture growth rates in the absence
5 of amino acid supplementation and in the presence of
 1×10^{-6} M 997 were reduced to 16% that of herbicide
free cultures. Supplementation with the branched chain
amino acids resulted in growth rates averaging 91% that of
control cultures, demonstrating an almost complete
10 alleviation of herbicide induced growth inhibition.

Experiments were conducted to determine the
optimum supplementation levels of each of the three amino
acids to obtain maximum herbicide protection and for
15 gaining insight into the site of action of the compound.
The most effective combination of concentrations employed
was 1 mM of each of the three amino acids. One mM each of
leucine and valine appeared to be the most effective
pairwise combination, and valine was most effective as a
20 lone supplement (See Table 2).

25

30

35

TABLE 2

5		Reversal of 997 Inhibition of Cell Suspension Culture Growth by Leucine, Valine and Isoleucine ¹					
		1,1,1	1,1,.3	1,1,.1	1,.3,1	1,.3,.3	1,.3,.1
	(-)	2.00	2.20	1.75	2.03	2.23	1.88
10	(+)	1.88	1.55	.93	1.45	1.40	.98
		1,.1,1	1,.1,.3	1,.1,.1	.3,1,1	.3,1,.3	.3,1,.1
	(-)	2.13	2.18	2.20	2.25	2.30	2.10
	(+)	1.28	1.38	1.03	1.30	1.30	1.08
15		.3,.3,1	.3,.3,.3	.3,.3,.1	.3,.1,1	.3,.1,.3	.3,.1,.1
	(-)	2.20	2.33	2.33	2.25	2.25	2.25
	(+)	1.08	1.18	.95	.80	.95	.70
		.1,1,1	.1,1,.3	.1,1,.1	.1,.3,1	.1,.3,.3	.1,.3,.1
	(-)	2.20	2.20	2.05	2.38	2.25	2.20
20	(+)	.60	.68	.83	.80	.90	.75
		.1,.1,1	.1,.1,.3	.1,.1,.1	0.3 val	0.3 leu	0.3 ile
	(-)	2.38	2.45	2.30	2.00	2.00	2.35
	(+)	.58	.68	.70	.48	.29	.33
25	No Supplementation (4 replicates)						
	(-)	2.36					
	(+)	0.30					

¹ Flasks were inoculated with 0.18 ml of cells per 10 ml media. Following growth for six days, the volume of cells per 10 ml media was quantitated. Numbers separated with commas refer to the mM concentrations of valine, leucine and isoleucine respectively. (+) refers to the presence of 1×10^6 M 997 and (-) to its absence. The values are the average of two replicates.

These results indicated that 997 induced growth inhibition and protein synthesis inhibition is a result of a decreased availability of the amino acids leucine, valine, and isoleucine. Additionally, the results implied that a major site of 997 action is associated with the leucine, valine, isoleucine biosynthetic pathway. It was further found that media supplementation with these three amino acids could reverse the growth inhibitory effects of 997 ranging in concentration from 1×10^{-8} to 1×10^{-3} M, indicating that this metabolic pathway contained the only significant site of action. (See FIG. 1.) Full reversal could not be obtained because of a toxic effect of the amino acids at high concentration.

Media supplementation with 1 mM each of leucine, isoleucine, and valine also reversed the growth inhibitory effects of two additional members of the imidazolinone family of chemicals structurally related to 997 (AC 252,214 [2-(5-isopropyl-5-methyl-4-oxo-2-imidazolin-2-yl)3-quinolinecarboxylic acid, hereinafter referred to as 214] and AC 263,499 [5-ethyl,2-(5-isopropyl-5-methyl-4-oxo-2-imidazolin-2-yl)nicotinic acid, hereinafter referred to as 499] (American Cyanamid Co., Princeton, NJ) on cell suspension cultures. The results demonstrated that the observations made are not unique to 997 and indicated that a whole family of chemicals share the same mechanism and site of action.

6.1.5. IDENTIFICATION OF THE SITE OF ACTION OF THE IMIDAZOLINONE HERBICIDES

30

Herbicide structural information and herbicide toxicity reversal studies with the branched chain amino acids indicated acetohydroxyacid synthase to be the herbicide site of action.

35

The primary site of action of AC 243,997 probably resides at some reaction in the biosynthesis of leucine and valine from pyruvate and isoleucine from alpha-ketobutyric acid. (See FIG. 2.) Four enzymes in this pathway are shared in the synthesis of each of the three amino acids. Since media supplementation with all three amino acids is required for maximum alleviation of herbicide activity, these four enzymes are the most likely sites for herbicide interaction. Furthermore, AC 243,997 bears some structural similarity to leucine and valine. The isopropyl and carboxyl moieties on the herbicidal molecule can exist in the same spatial configuration as the equivalent moieties on leucine or valine. The enzyme catalyzing the first reaction in the pathway from pyruvate, acetohydroxyacid synthase, appears to have two regulatory binding sites which recognize leucine and valine, respectively. The transaminase which catalyzes the last reaction in the pathway has a substrate binding site which presumably has some affinity for leucine, valine, and isoleucine, the products of the reaction. The above observations make these two enzymes the most reasonable candidates for further in vitro studies. The following paragraphs describe studies which showed that the imidazolinones affect the activity of ABAS in vitro.

Assays were conducted as a modification of the procedure of Smith et al. (1979, Molec. Gen. Genet. 169: 299-314). Plant cell culture material was ground with sand in a cold mortar and pestle in an equal quantity (w/v) of cell disruption media containing 0.05 M potassium phosphate (pH 7.0) 0.1 mM $MgSO_4$, and 0.5 mM dithiothreitol. The preparation was centrifuged at 11,000 x g for 20 minutes and the supernatant assayed for ABAS activity. Assays were conducted in 1-ml volumes containing 0.05 M potassium phosphate (pH 8.0), 40 mM

pyruvate, 5 mM $MgCl_2$, 0.1 mM thiamine pyrophosphate, 0.2 ml cell extract, and various concentrations of the herbicide of interest. The reaction mixtures were incubated at 30°C for 75 minutes and the reaction terminated with the addition of 0.1 ml of 50% (v/v) H_2SO_4 . Following further incubation at 40°C for 20 minutes, the acetoin formed from acetolactate by acidification was quantitated using the method of Westerfield (1945, J. Biol. Chem. 161: 495). To 0.2 ml of reaction mixture, 0.8 ml of water, 0.5 ml of 0.5% (w/v) creatine, and 0.5 ml of 5% (w/v) naphthol in 10% (w/v) NaOH was added. After mixing, solutions were incubated at room temperature for one hour. The development of pink color was quantitated by measuring light absorbance at 530 nm.

AHAS activity from maize cell cultures was found to be significantly inhibited by 997. Enzyme activity was reduced 50% by 8×10^{-6} M 997. AC 252,214 and AC 263,499 also inhibited the enzyme activity. This data strongly indicated that AHAS was the primary site of phytotoxic activity of the imidazolinone family of herbicides.

Selection and characterization of a mutant with an altered AHAS that conferred herbicide resistance at the cellular level are described in the following subsections.

6.2. CHARACTERIZATION OF THE EFFECTS OF THE IMIDAZOLINONE HERBICIDES AND STRATEGY FOR SELECTION OF HERBICIDE RESISTANT MAIZE CELLS

6.2.1. INITIATION AND MAINTENANCE OF MAIZE CELL CULTURES WHICH RETAIN PLANT REGENERATION CAPACITY

Frable, embryogenic maize callus cultures were initiated from hybrid immature embryos produced by

pollination of inbred line A188 plants (University of Minnesota, Crop Improvement Association) with pollen of Inbred line B73 plants (Iowa State University). Ears were harvested when embryos had reached a length of 1.5 to 2.0 mm. The whole ear was surface sterilized in 50 v/v commercial bleach (2.63% w/v sodium hypochlorite) for 20 minutes at room temperature. The ears were then washed with sterile distilled, deionized water. Immature embryos were aseptically isolated and placed on nutrient agar initiation/maintenance media with the root/shoot axis exposed to the media. Initiation/maintenance media consisted of N6 basal media (Chih-ching in Proceedings of Symposium on Plant Tissue Culture, May 25-30, 1978, Science Press, Peking, pp. 43-50) with 2% (w/v) sucrose, 1.5 mg per liter 2,4-D, 6 mM proline, and 0.9% (w/v) agar.

The immature embryos were incubated at 26°C under dim light. Cell proliferations from the scutellum of the immature embryos were evaluated for friable consistency and the presence of well defined somatic embryos. Tissue with this morphology was transferred to fresh media 10 to 14 days after the initial plating of the immature embryos. The tissue was then subcultured on a routine basis every 12 to 16 days. Sixty to eighty mg quantities of tissue were removed from pieces of tissue that had reached a size of approximately one gram and transferred to fresh media. Subculturing always involved careful visual monitoring to be sure that only tissue of the correct morphology was maintained. The presence of the somatic embryos ensured that the cultures would give rise to plants under the proper conditions.

6.2.2. CALLUS CULTURE GROWTH INHIBITION
BY IMIDAZOLINONE HERBICIDES

Since maize is generally grown in the field after
5 soybean, it seemed reasonable to carry out selections for
maize resistance to 214 (a soybean herbicide) rather than
997 (a total vegetative compound). Therefore the effects
of 214 were determined on callus growth. Quantities of
tissue averaging 90 mg in size were transferred to
10 nutrient agar maintenance media containing concentrations
of 214 ranging from 0.003 to 3 mg per liter. Eight tissue
pieces were placed on each plate with three plates per
herbicide concentration. After 14 days growth, the tissue
was weighed again. Inhibition of growth rate was found to
15 be half maximal in the 0.1 to 0.3 mg per liter range of
concentration of 214. Cell death was not observed to
occur over a 14 day period at any of the tested herbicide
concentrations. However, culture morphology changes did
occur at herbicide concentrations that gave more than half
20 maximal inhibition of growth rate. Especially obvious was
the loss of somatic embryo formation.

Growth inhibition studies carried out over
periods of time greater than 14 days, in which tissue was
25 grown either in the presence of herbicide on the same
plates for more than 14 days or was transferred to fresh
plates containing the same herbicide concentration, showed
half maximal growth inhibitory effects at smaller
herbicide concentrations (3 to 10 fold). Therefore, the
30 growth inhibitory effects of the herbicide increased with
time as well as with increased quantities of herbicide in
the media. This observation demonstrated that the
herbicide was very persistent, i.e., it was probably not
metabolized to any great extent and accumulated in the
35 tissue.

6.2.3. REVERSIBLE EFFECTS OF 214

To determine whether the effects of 214 were reversible, Black Mexican Sweet Corn cells growing in liquid suspension culture were exposed to 0.03 mg per liter 214 for various periods of time up to 72 hours. Growth was completely inhibited. The cells were then washed with herbicide-free media and the growth rate evaluated again in the absence of herbicide. Cells were found to rapidly return to an uninhibited growth rate. During the herbicide exposure, cells did not divide but remained viable and resumed divisions once the herbicide was removed. The reversible nature of the herbicide induced growth inhibition indicated that continuous herbicide exposure should be used in the selection procedure as opposed to intermittent exposures of a given duration.

6.3. SELECTION AND CHARACTERIZATION OF AN HERBICIDE RESISTANT CELL LINE

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The selection protocol used to identify and isolate herbicide resistant cells was formulated to take into account that: (1) the effects of the herbicides were reversible; (2) the effects of the herbicides increased over time; and (3) high herbicide concentrations adversely affected somatic embryo development and potentially plant regeneration capacity. Therefore, the procedure involved exposing tissue continuously to low herbicide concentrations over several subculture intervals and keeping careful records of the growth of all tissue in the selection. In this way the herbicide was allowed to take effect slowly with continuous selection pressure permitting herbicide tolerant cells to accumulate over time and yet not affect the potential for plant

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regeneration. This procedure allowed for the selection of cells with even small levels of herbicide tolerance (2 to 3 fold in herbicide concentration).

5 6.3.1. SELECTION OF AN HERBICIDE RESISTANT CELL LINE

Many selections were carried out utilizing the selection protocol described supra. The selection of one such herbicide resistant line that was identified and
10 characterized is described below in detail.

Approximately thirteen grams of vigorously growing maize callus tissue was transferred to maintenance media in petri plates containing 0.03 mg per liter of
15 herbicide 214. Thirty plates were prepared and 80-90 mg of tissue were plated in 5 areas on each plate. The herbicide level was chosen from growth inhibition studies to provide less than 20% to 40% growth inhibition during the first two weeks of herbicide exposure.

20

After 10 days the tissue had increased in mass over four fold. Eighty to 90 mg pieces of tissue showing vigorous growth rate and retention of embryogenic morphology (i.e., presence of somatic embryos) were
25 subcultured on fresh media containing 0.03 mg per liter 214. Thirty plates containing an average of seven pieces of tissue per plate were prepared. Each piece of tissue was labeled and became the progenitor of a "line." A complete geneology was maintained and recorded for future
30 subcultures. Subsequent subculture intervals ranged from 15 to 30 days depending on overall growth of the callus tissue.

For each transfer all tissue showing growth and
35 somatic embryo forming ability were placed on fresh media,

and the number of uniform sized callus pieces within a given "line" was recorded. During the course of the selection process, the total number of lines decreased as the herbicide mediated growth inhibition became more
5 intense. Many lines however, increased in size. At the end of the fifth selection cycle, the level of 214 was raised to 0.1 mg per liter to increase the selection pressure.

10 Between the fifth and sixth selection cycles, the amount of tissue in line XA17 began to increase significantly. The tissue showing the increased growth rate had a morphology different from the parent material from which the selection was initiated. Cells were
15 smaller on the average and more cytoplasmically dense, giving the tissue a yellow hue. The tissue continued to grow rapidly over subsequent subculture intervals until the vast majority of the tissue in the selection was identified as line XA17. At the seventh subculture,
20 tissue of line XA17 was removed from the selection for the purpose of characterization. This stock tissue was maintained and accumulated on maintenance media described supra in Section 6.2.1. containing 0.1 mg per liter 214.

25 6.3.2. CHARACTERIZATION OF MAIZE CELL LINE XA17

The resistant cell line was characterized to evaluate: (1) the magnitude of the resistance; (2) the chemical spectrum of the resistance; and (3) the
30 biochemical basis for the resistance.

Initially, the maximum level of 214 in which line XA17 would grow stably was determined. Stock tissue which had been maintained in the presence of 0.1 mg per liter
35 214 was transferred to media containing 214 levels ranging

up to 30 mg per liter. Tissue was maintained for several subculture intervals and the growth rate evaluated. Line XA17 tissue grew normally at a maximum level of 214 of 3.0 mg per liter as compared with a maximum of 0.01 mg per 5 liter 214 for unselected tissue.

Two week growth inhibition studies were carried out (as described supra) with 214, 997, 499, and chlorsulfuron. The levels of 214, 997 and 499 giving a 10 50% growth inhibition were found to be 30 mg per liter for 214, and higher than 30 mg per liter for 997 and 499 as compared with levels of 0.1 to 0.3 mg per liter for unselected tissue. The growth rate of line XA17 tissue was 50% inhibited by 10 mg per liter chlorsulfuron 15 2-chloro-N-[(4-methoxy-6-methyl-1,3,5-triazin-2-yl) aminocarbonyl] benzenesulfonamide, also known as chlorsulfuron, DPX 4189, E.I. DuPont de Nemours and Company, Wilmington, DE) as compared with 0.01 mg per liter for unselected tissue. Although line XA17 was 20 selected for resistance to 214, it expressed at least a three hundred fold enhancement of resistance (on a concentration basis) to the other compounds tested.

6.3.3. AHAS ACTIVITY OF MAIZE LINE XA17

25

Assays were conducted to determine if acetohydroxyacid synthase extracted from line XA17 tissue was altered with respect to herbicide sensitivity. Such a change would signify that an alteration of this enzyme conferred the herbicide resistance. Assays were conducted 30 as described supra. AHAS activity of control tissue was 50% inhibited by 997, 214 and 499 in the concentration range of 1×10^{-6} to 1×10^{-5} M. AHAS activity of line XA17 tissue was 50% inhibited by 997, 214 and 499 in 35 the 3×10^{-3} to 3×10^{-2} M range of herbicide

concentration, indicating at least a 1000 fold change in herbicide sensitivity on a concentration basis. AHAS activity of control tissue was found to be 50% inhibited by 1×10^{-8} M chlorsulfuron compared with a 50% inhibition of line XA17 AHAS at 3×10^{-7} M.

The results demonstrate that AHAS is the site of action of the herbicides tested, and furthermore, resistance was conferred by an alteration of the site of action. Since line XA17 expresses resistance at the cellular level by virtue of having an altered site of action, the herbicide tolerance is likely to extend to many compounds that act at the same sites on AHAS, including all members of the imidazolinone and sulfonamide families of herbicides.

The imidazolinone and sulfonamide families of herbicides are structurally dissimilar. Although they both inhibit AHAS activity in plants they are likely to be interacting with different sites on the enzyme. If they interact with different sites, then the alteration in XA17 AHAS which gave rise to imidazolinone resistance is apparently substantial enough to affect both sites. The AHAS alteration of XA17 tissue may also affect the binding of compounds other than the imidazolinones or sulfonamides which interact with the enzyme.

Whether the above explanation of sites of action affected by the herbicides is correct or not, the fact remains that callus tissue and regenerated plants of line XA17 express substantial amounts of tolerance to members of both families of herbicides compared with other maize cell cultures or plants.

6.4. PLANT REGENERATION AND PRODUCTION OF SEED

6.4.1. PLANT REGENERATION PROTOCOL

5 Eighty to 90 mg quantities of maize callus tissue
were transferred to embryo and shoot development
(regeneration) media in petri plates. The media consisted
of MS basal media supplemented with 0.1 mg per liter 2,4-D
and 1×10^{-7} M abscissic acid. 0.25% Gelrite (Kelco
10 Co., San Diego, CA) was used as a solid support in place
of agar. The tissue was incubated in the dark at 26°C for
one week. The plates were then transferred to a light
regime at 26°C (approximately 300 footcandles, soft white
fluorescent) on a cycle of 14 hours light, 10 hours dark.
15 Developing plants reaching a size of 1-3 cm were then
transferred to flasks or jars, containing MS media with no
supplementation, for further development. When plants
reached the two to three leaf stage, they were moved to
pots containing vermiculite and exposed to light at 2600
20 footcandles. These plants were watered with 20% v/v MS
media for one week and then with water until they grew
stably. The plants were then transferred to soil for
growth to maturity.

25 6.4.2. HERBICIDE INHIBITION OF PLANT REGENERATION

The effects of various concentrations of 214 on
plant regeneration (embryo and shoot development) were
evaluated. Unselected culture tissue was transferred to
30 regeneration media containing various concentrations of
214 ranging from 0.01 to 0.3 mg per liter. The tissue was
then put through the first three weeks of the plant
regeneration protocol described supra. The culture was
scored after this time for the production of well defined
35 shoots.

Plant regeneration was 50% inhibited by 214 in the concentration range of 0.03 to 0.1 mg per liter based on the number of shoots formed. 0.3 mg per liter 214 completely inhibited somatic embryo development and shoot formation.

6.4.3. EXPRESSION OF HERBICIDE RESISTANCE IN PLANTS REGENERATED FROM LINE XA17 CALLUS TISSUE

10 Traits which are selected for and expressed in undifferentiated cell cultures are not necessarily expressed in the differentiated organs and tissues of a developing plant. Therefore, it was necessary to examine plants regenerated from line XA17 callus tissue in the
15 presence of normally inhibitory concentrations of 214. Line XA17 tissue was transferred to regeneration media containing 0.1, 0.3, and 3.0 mg per liter 214. Normally developing plantlets were obtained at all three herbicide levels, demonstrating that herbicide resistance was
20 expressed during plant development.

6.4.4. REGENERATION OF MATURE PLANTS AND PRODUCTION OF SEED

25 Mature plants were regenerated from line XA17 callus tissue that had either been removed from herbicide exposure for at least three subculture cycles or had been continuously exposed to 0.1, 0.3 or 3.0 mg per liter 214 on maintenance media. Plant regeneration frequency in all
30 instances was approximately 1 plant per 100 g cell culture. Plants were grown to maturity, and the pollen obtained was used to fertilize plants of inbred line B73. Conversely, pollen from B73 plants was crossed to line XA17 regenerated plants. The seed obtained was found to
35 germinate under normal planting conditions. The level of

expression and genetics of the resistance trait in the progeny of the regenerated plants is presently being studied.

5 6.4.5. EXPRESSION OF HERBICIDE RESISTANCE IN
 PROGENY OF REGENERATED PLANTS

Seedling assays of progeny obtained from plants regenerated from cell line XA17 were performed as described
10 below to determine whether the herbicide resistance selected in culture was inherited and expressed in plants following a sexual cycle.

An XA17 regenerated plant with a normal ear was
15 pollinated with pollen obtained from a herbicide sensitive inbred B73 plant grown from seed. Seeds were obtained and harvested approximately forty days post pollination.

Seeds to be used as controls in the heritability
20 assay were obtained from regenerated plants from an herbicide sensitive cell line of the same genetic background as line XA17. These regenerated plants were crossed with pollen obtained from a herbicide sensitive inbred B73 plant grown from seed. Seeds were harvested
25 approximately forty days after pollination.

Twenty control seeds and ten XA17 seeds were surface sterilized with 2.6% sodium hypochlorite for twenty minutes and then washed with sterile distilled
30 water. The seed was imbibed overnight in sterile water containing a small quantity of the fungicide, captan (N-trichloromethylthio-4-cyclohexene-1,2-dicarboximide). The mature embryos were then excised from the seed and
— plated scutellum side down on solid MS media without 2,4
35 D. Ten control seeds were plated on media containing no

214 and ten were plated in the presence of 6×10^{-8} M 214 which had been incorporated into the medium after autoclaving but before gelling. All ten XA17 seeds were plated in the presence of 6×10^{-8} M 214. Seedlings were 5 evaluated seven days after plating.

Control seedlings in the absence of 214 grew normally throughout the seven day period attaining an average shoot length of 11.8 cm (FIG. 3) and an average 10 root length of 8.1 cm (FIG. 4). Growth inhibition of control seedlings in the presence of 214 was first observed on day three. No further growth took place in subsequent days. The average shoot length and root length attained were 1.5 and 1.1 cm respectively.

15

Of the ten XA17 seedlings grown in the presence of 214, the growth of five seedlings was inhibited to the same degree as control seedlings grown in the presence of 214. Five XA17 seedlings showed resistance to growth 20 inhibition by 214 (FIG. 3 and FIG. 4).

Root growth of the resistant seedlings was particularly striking. The roots penetrated the herbicide containing media and grew rapidly with substantial 25 branching and attained lengths of 6-7 cm. Roots of sensitive seedlings grew little (0-1 cm) and remained on the surface of the media. The data indicate that the herbicide resistance trait which was selected in culture is inherited as a dominant gene (or genes) and that it 30 provides a high level of resistance to 214 in plants.

Resistant and sensitive seedlings were transferred to pots for growth to maturity. The trait will be crossed from these plants into agronomically 35 important inbred lines for commercial development.

6.4.6. METHOD FOR OBTAINING UNIFORM
HERBICIDE RESISTANT SEED

Seed from herbicide resistant regenerated plants
5 obtained from crosses with herbicide sensitive plants
segregate one resistant to one sensitive for the herbicide
resistance trait. That is, half the seeds possess a
single gene encoding resistance and half the seeds have no
resistance. Seeds which uniformly express the trait and
10 are heterozygous or homozygous with respect to the
resistant acetohydroxyacid synthase gene may be obtained
by performing the following crosses and assays.

Seed obtained from herbicide resistant
15 regenerated plants which have been crossed with sensitive
plants are planted, grown to sexual maturity, and self
pollinated. Samples of seed harvested from the resulting
ears are assayed for herbicide resistance using a seedling
assay. Half the ears will have no resistant seed. The
20 other half will have seed expressing the resistance trait
with the trait segregating 1:2:1 (homozygous resistant:
heterozygous resistant: homozygous sensitive), i.e., 75%
of the seed carrying the resistance trait.

Seed from the ears possessing the resistance
25 trait are then grown and self pollinated. Seed from the
resulting ears are again assayed for resistance.
Twenty-five percent of these ears will have seed that
uniformly express herbicide resistance. These seed are
homozygous for the resistance trait and come from
30 homozygous plants.

Seed that are uniformly heterozygous for the
resistance trait may be obtained by growing plants from
the homozygous seed and using these plants as either male
35

or female parents in crosses with plants lacking the resistance gene. Progeny from these crosses are uniformly herbicide resistant and heterozygous for the resistance gene.

5

7. EXAMPLE

7.1. SELECTION OF ADDITIONAL HERBICIDE RESISTANT MAIZE CELL LINES

10

The following sections briefly describe the selection and partial characterization of two additional imidazolinone resistant maize cell lines, QJ22 and UV18.

15

7.1.2. SELECTION OF MAIZE CELL LINE QJ22

Line QJ22 was selected from maize cell culture using the protocol described in detail for the selection of line XA17 (Section 6.3.1) with the following significant differences: (1) maize callus tissue was selected using the herbicide 499; and (2) the tissue used to initiate the selection was obtained from a previously selected cell line, XA119. XA119 was identified in a selection for 214 tolerance and had been demonstrated to have an approximately three fold enhancement of tolerance of 214 at the cellular level.

The selection line QJ22 was initiated at a 499 concentration of 0.1 mg per liter. The selection was maintained through two very long subculture intervals covering a 12 week period of time. Line QJ22 was identified after 12 weeks of selection as a rapidly growing sector of tissue with good tissue morphology. QJ22 tissue was increased over the following weeks in the presence of 0.1 mg per liter 499. After 8 weeks of

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accumulation, the cell line was characterized and plant regeneration efforts initiated. Several shoots and small plants were obtained and are being grown to maturity.

5 7.1.3. SELECTION OF LINE UV18

Line UV18 was also selected from maize cell culture using the protocol described in detail for the selection of line XA17 with one important difference. The
10 tissue used to initiate the selection was subjected to irradiation with ultraviolet light before being exposed to 214.

For the ultraviolet light exposure, tissue was spread in a thin layer (1-3 mm) on maintenance media in
15 plastic petri dishes. These plates were then irradiated with UV light for periods of 0, 1, 2, 4, and 8 minutes in the absence of visible light. The ultraviolet light source was a Westinghouse model 782L-30 29W lamp (Westinghouse Electrical Corporation, Lamp Commercial
20 Division, Bloomfield, NJ) placed approximately seven inches from the tissue. The irradiated tissue was grown in the dark for two weeks. Surviving tissue was then transferred to maintenance media containing 0.1 mg per liter 214 to initiate selection for 214 resistance.

25 Line UV18 was identified from tissue which had obtained a 2 minute exposure to ultraviolet light. It could not be determined whether the alteration giving rise to 214 tolerance was a result of the ultraviolet radiation
30 or a spontaneous variation that occurred during the selection process.

The UV18 cell line was first observed after 25 weeks of selection comprising 8 subculture intervals. The
35 tissue in the selection procedure had been exposed to 0.1

mg per liter 214 for four subculture intervals followed by exposure to 0.3 mg per liter 214 for four subculture intervals. UV18 was identified during the ninth subculture as a rapidly growing sector of tissue with good morphology. The tissue grew as rapidly in the presence of 0.3 mg per liter 214 as did control tissue in the absence of herbicide.

The cell line was rapidly increased in the presence of 214 and characterized. Plants were regenerated in the presence of normally toxic levels of 214 (0.3 mg per liter) demonstrating the resistance trait was expressed in differentiated plant organs. Herbicide resistant plants regenerated from the UV18 cell culture are presently being grown to maturity.

7.1.4. CHARACTERIZATION OF LINES QJ22 AND UV18

The resistant callus tissues were characterized to determine the magnitude and spectrum of herbicide tolerance.

Callus tissue from lines QJ22 and UV18 was subjected to growth inhibition studies with a series of concentrations of the compounds 214, 997, 499, and chlorsulfuron. Table 3 describes the results of these studies. The numbers represent the fold increase in herbicide concentration that can be tolerated by each of the cell lines as compared to control tissue. The values were calculated based on the amount of herbicide that gave a 50% reduction in growth rate. In the instance where greater than 100 fold resistance is displayed, the tissue was completely tolerant of the highest concentration tested. UV18, in general, showed a tolerance of the herbicides similar to that of line XA17. QJ22 is

significantly different from UV18 and XA17, i.e., a much smaller degree of tolerance of 214 and essentially no tolerance of chlorsulfuron were observed.

5

TABLE 3

Callus Growth

10	<u>Selected Line</u>	<u>Fold Increase in Tolerance</u>			
		<u>997</u>	<u>214</u>	<u>499</u>	<u>Chlorsulfuron</u>
	QJ22	30-50	5	30-50	0
	UV18	100	100	100	300

15

7.1.5 HERBICIDE INHIBITION OF AHAS ACTIVITY
OF MAIZE LINES QJ22 AND UV18

Acetohydroxacid synthase activity was extracted from cell lines UV18 and QJ22 and assayed as described in detail for cell line XA17, supra. AHAS activity from both selected lines was more tolerant of 997 over a 100 fold range of 997 concentrations (3×10^{-6} to 3×10^{-4} M) than was AHAS activity from control cultures. The results suggest that UV18 and QJ22, like XA17, are herbicide resistant by virtue of having an altered herbicide site of action. Additionally, other mechanisms may be involved in imparting herbicide resistance in these cell lines.

25

8. DEPOSIT OF CELL LINES AND SEEDS

30

The following cell lines, as described herein have been deposited with the American Type Culture Collection, Rockville, Maryland, and have been assigned the following accession numbers:

35

<u>Cell Line</u>	<u>Accession Numbers</u>
XA17	40100
QJ22	40129
UV18	40128

5

Seeds derived from plants grown from cell line XA17 as described herein have been deposited with In Vitro International, Inc., Ann Arbor, Michigan and have been assigned IVI accession No. 10011.

10

The present invention is not to be limited in scope by the cell line or seeds deposited, since the deposited embodiments are intended as single illustrations of one aspect of the invention and any cell lines or seeds which are functionally equivalent are within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

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9,702 WHAT IS CLAIMED IS:

1. A maize plant wherein the growth and development of said plant is resistant to inhibition by a 2-(2-imidazolin-2-yl)pyridine herbicide, or a derivative thereof at levels which normally inhibit the growth and development of said maize plant; a maize plant wherein the growth and development of said plant is resistant to inhibition by a 2-(2-imidazolin-2-yl)quinoline herbicide, or a derivative thereof at levels which normally inhibit the growth and development of said maize plant; or a maize plant wherein the growth and development of said plant is resistant to inhibition by a herbicidal sulfonamide at levels which normally inhibit the growth and development of said maize plant.

2. The maize plant according to claim 1, wherein the herbicide is AC 263,499; wherein the herbicide is AC 243,997; wherein the herbicide is AC 252,214; or wherein the herbicide is a 2-chloro-N[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide.

3. The maize plant according to claim 1, wherein said plant contains a gene conferring resistance to said herbicide that does not naturally occur in said plant; wherein the gene is from a eukaryotic organism; wherein the gene is from a prokaryotic organism; or wherein the gene is made in whole or part by chemical or enzymatic synthetic methods.

4. The maize plant according to claims 1 or 2, wherein the plant contains an altered acetohydroxyacid synthase resistant to inhibition by said herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase.

5. The maize plant according to claims 3, wherein the gene conferring resistance encodes an altered acetohydroxyacid synthase resistant to inhibition by the herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said plant capable of expressing said gene.

6. An herbicide resistant maize plant derived from maize cell line XA17, deposited with the ATCC and assigned accession No. 40100, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize plant derived from maize cell line QJ22, deposited with the ATCC and assigned accession No. 40129, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize plant derived from maize cell line UV18, deposited with the ATCC and assigned accession NO. 40128, and mutants, recombinants and genetically engineered derivatives thereof; or an herbicide resistant maize plant derived from seeds deposited with In Vitro International, Inc. and assigned IVI assession No. i0011, and mutants, recombinants and genetically engineered derivatives thereof.

7. A plant containing a gene coding for an altered acetohydroxyacid synthase resistant to inhibition by an herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said plant capable of expressing said gene.

8. The plant according to claim 7, wherein the plant is a monocot; wherein the plant is a cereal crop; or wherein the plant is selected from a group consisting of maize, rice, wheat, barley, sorghum, oats, rye and millet.

9. The plant according to claim 8 wherein the herbicide is 2-(2-imidazolin-2-yl)pyridine, or a derivative thereof; wherein the herbicide is AC 263,499; wherein

the herbicide is AC 243,997; wherein the herbicide is 2-(2-imidazolin-2-yl)quinoline, or a derivative thereof; wherein the herbicide is AC 252,214; wherein the herbicide is a herbicidal sulfonamide; or wherein the herbicide is a 2-chloro-N-[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide.

10. A maize tissue culture wherein the growth of said tissue culture is resistant to inhibition by a 2-(2-imidazolin-2-yl)pyridine herbicide, or a derivative thereof at levels which normally inhibit the growth of said tissue culture; wherein the herbicide is AC 263,499; or wherein the herbicide is AC 243,997; a maize tissue culture wherein the growth of said tissue culture is resistant to inhibition by a 2-(2-imidazolin-2-yl)quinoline herbicide, or a derivative thereof at levels which normally inhibit the growth of said tissue culture; or wherein the herbicide is AC 252,214; or a maize tissue culture wherein the growth of said tissue culture is resistant to inhibition by a herbicidal sulfonamide at levels which normally inhibit the growth of said tissue culture; or wherein the herbicide is a 2-chloro-N[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)-aminocarbonyl] benzenesulfonamide.

11. The maize tissue culture according to claim 10, wherein said tissue culture contains a gene conferring resistance to said herbicide that does not naturally occur in said tissue culture; wherein the gene is from a eukaryotic organism; wherein the gene is from a prokaryotic organism; or wherein the gene is made in whole or part by chemical or enzymatic synthetic methods.

12. The maize tissue culture according to claim 10 wherein the tissue contains an altered acetohydroxyacid synthase resistant to inhibition by said herbicide at concentrations which normally inhibit the activity of an

unaltered acetohydroxyacid synthase.

5 13. The maize tissue culture according to claim 11, wherein the gene conferring resistance encodes an altered acetohydroxyacid synthase resistant to inhibition by the herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said tissue culture capable of expressing said gene.

10 14. An herbicide resistant maize tissue culture derived from maize cell line XA17, deposited with the ATCC and assigned accession No. 40100, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize tissue culture derived from
15 maize cell line QJ22, deposited with the ATCC and assigned accession No. 40129, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize tissue culture derived from maize cell line UV18, deposited with the ATCC and assigned accession No.
20 40128, and mutants, recombinants and genetically engineered derivatives thereof; or an herbicide resistant maize tissue culture derived from seeds deposited with In Vitro International, Inc. and assigned IVI assession No. 10011, and mutants, recombinants and genetically engineered derivatives thereof.

25 15. The plant tissue culture according to claim 14 wherein the herbicide is 2-(2-imidazolin-2-yl)pyridine, or a derivative thereof; wherein the herbicide is AC 263,499; wherein the herbicide is AC 243,997; wherein the herbicide
30 is 2-(2-imidazolin-2-yl)quinoline, or a derivative thereof; wherein the herbicide is AC 252,214; wherein the herbicide is a herbicidal sulfonamide; or wherein the herbicide is a 2-chloro-N-[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide.

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16. A plant tissue culture containing a gene cod-

ing for an altered acetohydroxyacid synthase resistant to inhibition by an herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said tissue culture capable of expressing said gene; wherein the tissue culture is derived from a monocot; wherein the tissue culture is derived from a cereal crop; or wherein the tissue culture is selected from a group consisting of maize, rice, wheat, barley, sorghum, oats, rye and millet.

10

17. The plant tissue culture according to claim 16 wherein said tissue contains a gene conferring resistance to said herbicide that does not naturally occur in said tissue; wherein the gene is from a eukaryotic organism; wherein the gene is from a prokaryotic organism; or wherein the gene is made in whole or part by chemical or enzymatic synthetic methods.

15

18. A maize seed from which a maize plant can be grown wherein the growth and development of said plant is resistant to inhibition by a 2-(2-imidazolin-2-yl)pyridine herbicide, or a derivative thereof at levels which normally inhibit the growth and development of said plant; wherein the herbicide is AC 263,499; or wherein the herbicide is AC 243,997; a maize seed from which a maize plant can be grown wherein the growth and development of said plant is resistant to inhibition by a 2-(2-imidazolin-2-yl)quinoline herbicide, or a derivative thereof at levels which normally inhibit the growth and development of said plant; or wherein the herbicide is AC 252,214; or a maize seed from which a maize plant can be grown wherein the growth and development of said plant is resistant to inhibition by a herbicidal sulfonamide at levels which normally inhibit the growth and development of said plant; or wherein the herbicide is a 2-chloro-N[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide.

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19. The maize seed according to claim 18, wherein said seed contains a gene conferring resistance to said herbicide that does not naturally occur in said seed; wherein the gene is from a eukaryotic organism; wherein the gene is from a prokaryotic organism; or wherein the gene is made in whole or part by chemical or enzymatic synthetic methods.

20. The maize seed according to claim 18 wherein the seed contains an altered acetohydroxyacid synthase resistant to inhibition by an herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase.

21. A maize seed according to claim 19, wherein the gene conferring resistance encodes an altered acetohydroxyacid synthase resistant to inhibition by the herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said plant capable of expressing said gene.

22. An herbicide resistant maize seed derived from maize cell line XA17, deposited with the ATCC and assigned accession No. 40100, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize seed derived from maize cell line QJ22, deposited with the ATCC and assigned accession No. 40129, and mutants, recombinants and genetically engineered derivatives thereof; an herbicide resistant maize seed derived from maize cell line UV18, deposited with the ATCC and assigned accession No. 40128, and mutants, recombinants and genetically engineered derivatives thereof; or an herbicide resistant seed deposited with In Vitro International, Inc. and assigned IVI assession No. 10011, and mutants, recombinants and genetically engineered derivatives thereof.

23. A seed containing a gene coding for an altered acetohydroxyacid synthase resistant to inhibition by an herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase, said plant capable of expressing said gene; wherein the seed is derived from a monocot; wherein the seed is derived from a cereal crop; wherein the seed is selected from a group consisting of maize, rice, wheat, barley, sorghum, oats, rye and millet; wherein the herbicide is 2-(2-imidazolin-2-yl)pyridine, or a derivative thereof; wherein the herbicide is AC 263,499; wherein the herbicide is AC 243,997; wherein the herbicide is 2-(2-imidazolin-2-yl)quinoline, or a derivative thereof; wherein the herbicide is AC 252,214; wherein the herbicide is a herbicidal sulfonamide; or wherein the herbicide is a 2-chloro-N-[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)aminocarbonyl] benzenesulfonamide.

24. A gene coding for an altered plant acetohydroxyacid synthase resistant to inhibition by a herbicide at concentrations which normally inhibit the activity of an unaltered acetohydroxyacid synthase.

25. A method for producing an herbicide resistant plant, plant tissue or plant seed comprising the steps of:

(a) initiating a plant tissue culture derived from immature embryos sterily obtained from a plant and plating said embryos on an initiation/maintenance media;

(b) growing a plant tissue culture derived from said embryos in the presence of an herbicide at a level which provides for less than from about 20% to about 40% growth inhibition during the first two weeks of said herbicide exposure;

(c) subculturing the plant tissue culture of step (b);

5 (d) identifying a stably herbicide resistant plant cell line from step (c) which grows at a rate comparable to that of the plant tissue culture in the absence of said herbicide; and

10 (e) regenerating a plant and producing a seed from said stably herbicide resistant plant cell line in the presence of a level of herbicide that exceeds by at least three fold the level of herbicide that would normally inhibit the growth of a regenerating maize plant.

15 26. The method according to claim 25, wherein the plant tissue culture in step (b) is first irradiated using ultraviolet light.

20 27. The method according to claim 26, wherein the herbicide comprises a 2-(2-imidazolin-2-yl)pyridine herbicide; wherein the herbicide comprises a 2-(2-imidazolin-2-yl)quinoline herbicide; wherein the herbicide comprises a herbicidal sulfonamide; wherein the plant comprises a
25 monocot; or wherein the plant comprises a cereal crop.

28. A method for producing an herbicide resistant maize plant, maize tissue or maize seed comprising the steps of:

30 (a) initiating a maize plant tissue culture derived from immature embryos sterily obtained from a maize plant and plating said embryos on an initiation/maintenance media;

35 (b) growing a maize plant tissue culture derived from said embryos in the presence of an

herbicide at a level which provides for less than from about 20% to about 40% growth inhibition during the first two weeks of said herbicide exposure;

5

(c) subculturing the maize plant tissue culture of step (b);

10

(d) identifying a stably herbicide resistant maize plant cell line from step (c) which grows at a rate comparable to that of the maize plant tissue culture in the absence of said herbicide; and

15

20

(e) regenerating a maize plant and producing a seed from said stably herbicide resistant maize plant cell line in the presence of a level of herbicide that exceeds by at least three fold the level of herbicide that would normally inhibit the growth of a regenerating maize plant.

25

29. The method according to claim 28, wherein the herbicide comprises a 2-(2-imidazolin-2-yl)pyridine herbicide; wherein the herbicide comprises a 2-(2-imidazolin-2-yl)quinoline herbicide; or wherein the herbicide comprises a herbicidal sulfonamide.

30

30. An herbicide resistant plant, plant tissue or plant seed produced by a process comprising the steps of:

35

(a) initiating a plant tissue culture derived from immature embryos sterilely obtained from a plant and plating said embryos on an initiation/maintenance media;

5 (b) growing a plant tissue culture derived from said embryos in the presence of an herbicide at a level which provides for less than from about 20% to about 40% growth inhibition during the first two weeks of said herbicide exposure;

10 (c) subculturing the plant tissue culture of step (b);

15 (d) identifying a stably herbicide resistant plant cell line from step (c) which grows at a rate comparable to that of the plant tissue culture in the absence of said herbicide; and

20 (e) regenerating a plant and producing a seed from said stably herbicide resistant plant cell line in the presence of a level of herbicide that exceeds by a least three fold the level of herbicide that would normally inhibit the growth of a regenerating plant.

25 31. The herbicide resistant plant, plant tissue or plant seed of claim 30, wherein the herbicide is a 2-(2-imidazolin-2-yl)pyridine herbicide; wherein the herbicide is a 2-(2-imidazolin-2-yl)quinoline herbicide; wherein the herbicide is a herbicidal sulfonamide; wherein the plant is a monocot; or wherein the plant is a cereal crop.

30 32. An herbicide resistant maize plant, maize tissue or maize seed produced by a process comprising the steps of:

35 (a) initiating a maize plant tissue culture derived from immature embryos sterilely obtained from a maize plant and plating said embryos on an initiation/maintenance media;

(b) growing a maize plant tissue culture derived from said embryos in the presence of an herbicide at a level which provides for less than from about 20% to about 40% growth inhibition during the first two weeks of said herbicide exposure;

(c) subculturing the maize plant tissue culture of step (b);

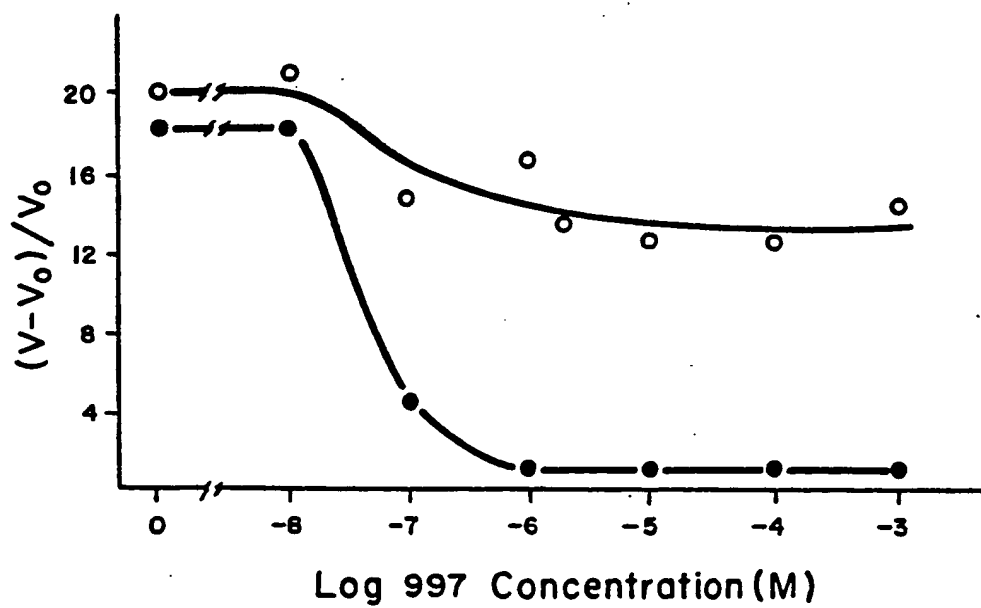
(d) identifying a stably herbicide resistant maize plant cell line from step (c) which grows at a rate comparable to that of the maize plant tissue culture in the absence of said herbicide; and

(e) regenerating a maize plant and producing a seed from said stably herbicide resistant maize plant cell line in the presence of a level of herbicide that exceeds by at least three fold the level of herbicide that would normally inhibit the growth of a regenerating maize plant.

33. The herbicide resistant maize plant, maize plant tissue or maize plant seed of claim 32, wherein the herbicide is a 2-(2-imidazolin-2-yl)pyridine herbicide; wherein the herbicide is a 2-(2-imidazolin-2-yl)quinoline herbicide; or wherein the herbicide is a herbicidal sulfonamide.

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FIG. 1



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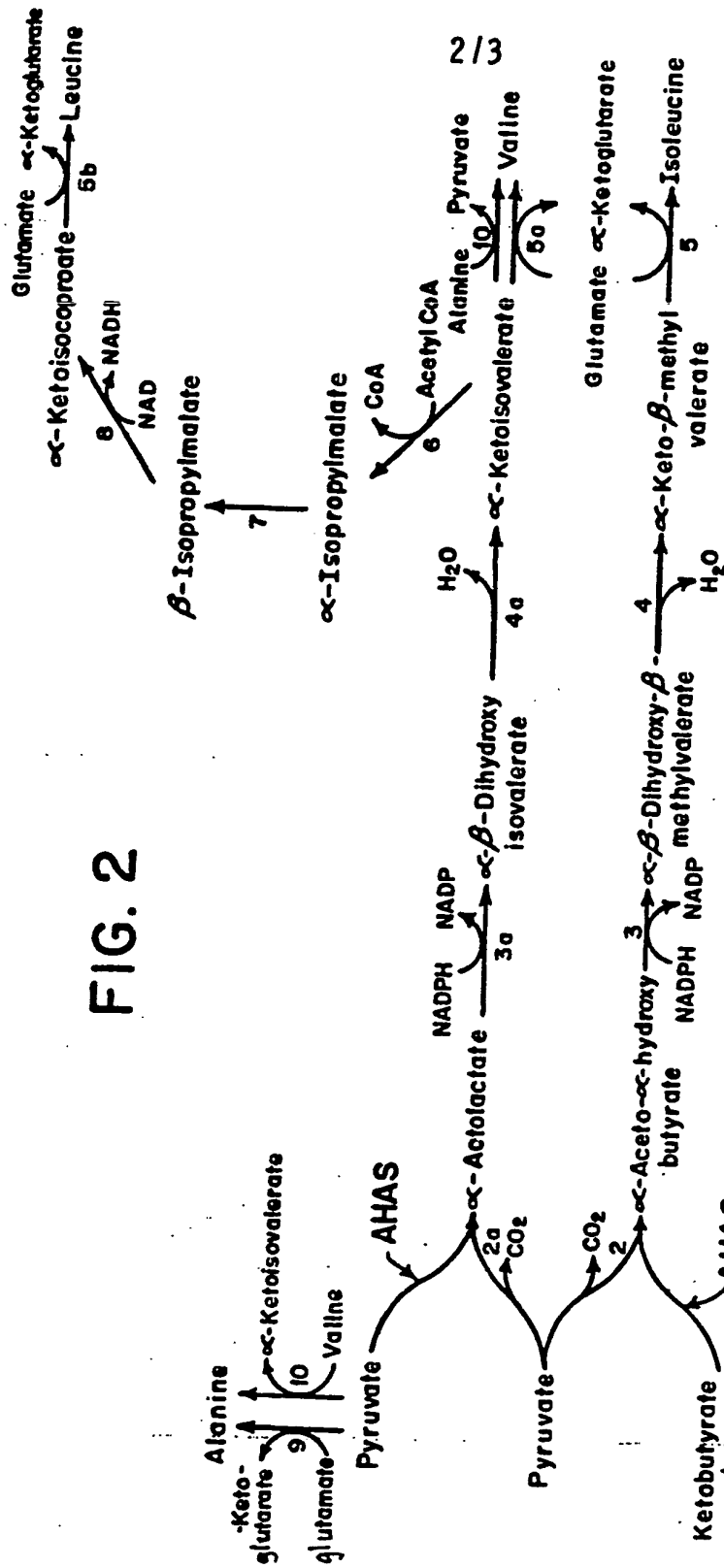


FIG. 2

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FIG. 3

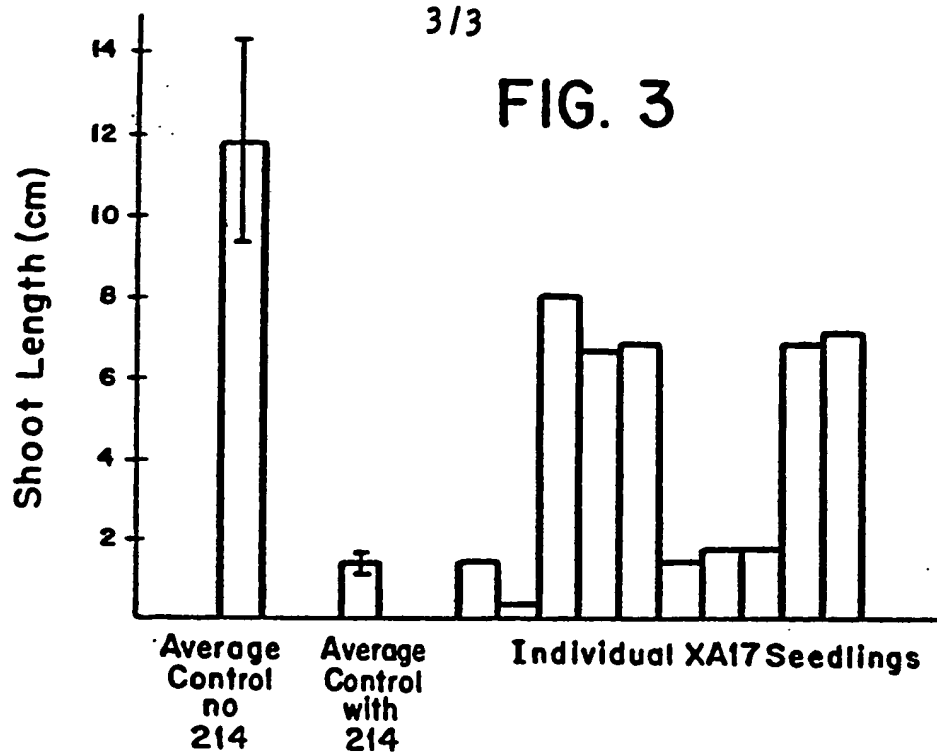
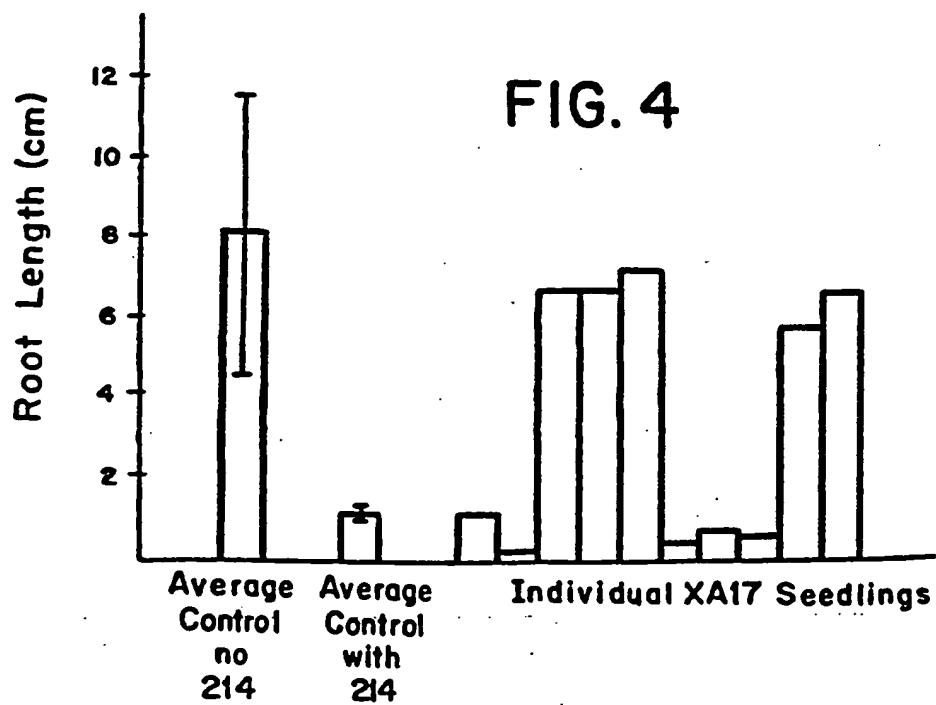


FIG. 4



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